

LITTER/FECAL FLOTATION PROTOCOL

David D. Frame, DVM, DACPV

Collection

1. Grab small handfuls of litter from various locations in the building, for example under waterers, around feeders, from middle of building, etc. Collect at least five representative samples. Or collect at least ten recently deposited fecal droppings. Collect them in a plastic ziplock bag, knead and mix thoroughly. Make sure the composite sample is well-mixed and homogenous.
2. Label bag with date of collection and location ID.
3. Store sealed bag under refrigeration if not being tested soon after collection.

Sugar Solution (1.27 specific gravity)

Water	100 ml
Table sugar.	128 g

Mix and heat until all sugar is dissolved. Store in a closed container.

Procedure

1. Weigh out 3 g (or use about ½ teaspoon) of the composite sample.
2. Add about 20 mL of the sugar solution to the 3 g sample.
3. Mix and strain through kitchen strainer.
4. Pour strained liquid into a 15 mL tube (e.g. plastic blood tubes with snap tops), or use an Ovassay container.
5. Using an eyedropper, carefully add enough sugar solution to finish filling up the tube until it forms a convex surface at top of tube.
6. Gently place a coverslip over this convex surface being careful not to get too many bubbles between the coverslip and liquid interface.
7. Let the tube with coverslip rest upright for 30 minutes.
8. After 30 minutes, gently lift coverslip from top of tube so as to not disturb the drop of accumulated debris, place on clean glass microscope slide, and read.